

The effect of FeCl₃ on the accumulation of gallium-67 into inflammatory and normal tissues

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The effect of FeCl₃ on the uptake of ⁶⁷Ga by inflammatory and normal tissues was studied to clarify the role of transferrin in ⁶⁷Ga uptake by inflammatory tissue. The administration of FeCl₃ 5 min before the injection of ⁶⁷Ga decreased the uptake of ⁶⁷Ga by liver and spleen, but had little effect on the uptake of ⁶⁷Ga by the inflammatory tissue. These results suggest that ⁶⁷Ga is taken up by normal tissues in a transferrin-bound form but in an unbound form by inflammatory tissue. On the other hand, when FeCl₃ was simultaneously injected with ⁶⁷Ga, the uptake of ⁶⁷Ga by liver and spleen was markedly increased but the uptake by inflammatory tissue was decreased.

Key words: ⁶⁷Ga uptake, Inflammation, FeCl₃

INTRODUCTION

GALLIUM-67 has been used for the detection of various tumors¹ and acute and chronic inflammation.^{2,3} Many hypotheses concerning the mechanism of the uptake of ⁶⁷Ga by tumors and inflammatory lesions⁴⁻⁵ have been proposed. Tsan et al.⁸ and Vallabhajosula et al.⁹ reported that ⁶⁷Ga was almost all bound to transferrin, iron-binding protein, in blood. It was reported that transferrin played a major role for the mechanism of the uptake of ⁶⁷Ga by tumors.¹⁰⁻¹³ On the other hand, it was shown that transferrin transports ⁶⁷Ga to the tumor tissue but that ⁶⁷Ga is taken up by tumor cells in an unbound form.^{7,14-16} The involvement of transferrin in the uptake of ⁶⁷Ga by inflammatory tissue has not, however, been conclusively demonstrated. We have recently demonstrated that ⁶⁷Ga densely accumulates in granuloma tissue, that is inflammatory tissue produced by turpentine oil.¹⁷ In the present study using FeCl₃, we have investigated whether or not transferrin is involved in the uptake of ⁶⁷Ga by the inflammatory tissue.

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MATERIALS AND METHODS

Animals

Male Wistar rats weighing 150-200 g were purchased from Shizuoka Laboratory Animal Center (Hamamatsu, Japan), and were housed in wire mesh cages at a room temperature of 23±1°C and in relative humidity of 55±5%.

Production of inflammatory tissue

The production of inflammatory tissue, granuloma, was carried out by the same method described in the previous report¹⁷ as follows. A paper pellet (size 8 mm) was dipped in turpentine oil and implanted bilaterally in the subcutaneous tissues of the abdomen in each animal.

⁶⁷Ga solution

Gallium-67 citrate (kindly supplied by Nihon Medipharma Co., Ltd., Takarazuka, Japan) was diluted with saline to 185 kBq (5 μCi)/ml.

Administration of ⁶⁷Ga

Each rat, at 6 days after the administration of turpentine oil, was intravenously injected with ⁶⁷Ga in a dose of 37 kBq (200 μl).

Administration of FeCl₃

Each rat, at 6 days after the administration of turpentine oil, was intravenously injected with FeCl₃ (6.25-25 μmole/ml saline) in a dose of 100 μl 5 min before or 30 min after the administration of ⁶⁷Ga.

The simultaneous administration of FeCl_3 and ^{67}Ga was carried out by the intravenous injection of FeCl_3 saline solution ($12.5 \mu\text{mole/ml}$) containing ^{67}Ga (185 kBq) in a dose of $200 \mu\text{l}$.

Removal of granuloma and other tissues

At 4 hrs after the administration of ^{67}Ga , rats were anesthetized with urethane (1.5 g/kg , i.p.) and immediately perfused with cold saline. The inflammatory lesion and other tissues were then removed. The granuloma tissue was obtained from the inflammatory lesion with complete removal of the implanted paper pellet and abscess.

Determination of radioactivity

Radioactivity of various tissues removed was determined with a well-type NaI-scintillation counter (Aloka, ARC-300). The uptake ratios of ^{67}Ga in various tissues were expressed in the following formula: Uptake ratio = sample radioactivity (cpm) / sample weight (g) / total radioactivity administered (cpm) / body weight of rat (g).

RESULTS

The effect of various amounts of FeCl_3 on the uptake of ^{67}Ga by inflammatory and normal tissues

To investigate the effect of FeCl_3 on the retention of ^{67}Ga in the blood and ^{67}Ga uptake by inflammatory and normal tissues, *in vivo* experiments were performed (Fig. 1). FeCl_3 (0.625 , 1.25 , and $2.5 \mu\text{mole/rat}$) was administered 5 min before the injection of ^{67}Ga and the blood, urine, inflammatory and other tissues were removed 4 hrs later. FeCl_3 dose dependently reduced ^{67}Ga retention in the blood. Similarly the uptake of ^{67}Ga by the liver and spleen was dose dependently reduced. In contrast to this,

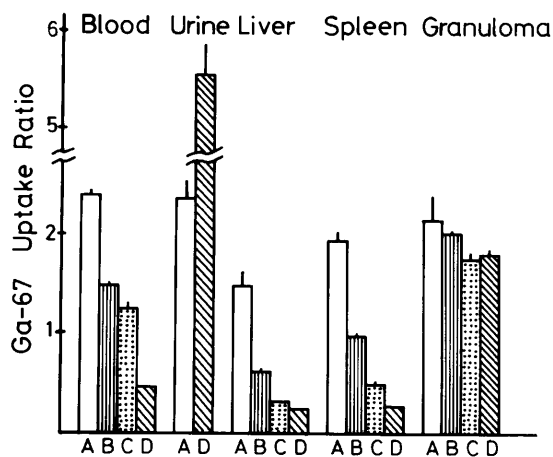


Fig. 1 Effect of various amounts of FeCl_3 on the uptake of ^{67}Ga by several tissues. A: ^{67}Ga alone. B: 0.625 , C: 1.25 and D: $2.5 \mu\text{mole/rat}$ of FeCl_3 were administered 5 min before the injection of ^{67}Ga . Each point represents the mean and SEM for five rats.

FeCl_3 ($2.5 \mu\text{mole/rat}$) markedly increased the excretion of ^{67}Ga into the urine. On the other hand, FeCl_3 had little effect on the uptake of ^{67}Ga into the inflammatory tissue, granuloma.

The effect of the time of FeCl_3 administration on the uptake of ^{67}Ga in inflammatory and normal tissues

The administration of FeCl_3 ($2.5 \mu\text{mole/rat}$) 30 min after the injection of ^{67}Ga decreased the uptake ratios of ^{67}Ga into the blood more than that 5 min before (Fig. 2). On the other hand, the administration of FeCl_3 30 min after the injection of ^{67}Ga increased the uptake of ^{67}Ga by the liver and spleen more than 5 min before. But in all cases, the blood, liver and spleen uptake ratios of ^{67}Ga were greatly decreased by the administration of FeCl_3 as compared with ^{67}Ga alone. On the other hand, concerning the uptake ratio of ^{67}Ga into granuloma, there is no difference between the administration of FeCl_3 5 min before and 30 min after the injection of ^{67}Ga , although the uptake ratios in both cases were slightly decreased by the administration of FeCl_3 .

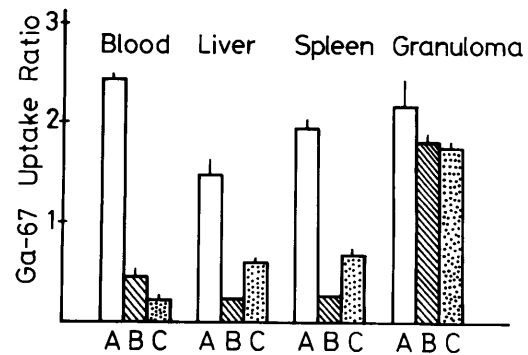


Fig. 2 Effect of the time of FeCl_3 administration on the uptake of ^{67}Ga by several tissues. A: ^{67}Ga alone. FeCl_3 ($2.5 \mu\text{mole/rat}$) was administered 5 min before (B) and 30 min after (C) the injection of ^{67}Ga . Each point represents the mean and SEM for five rats.

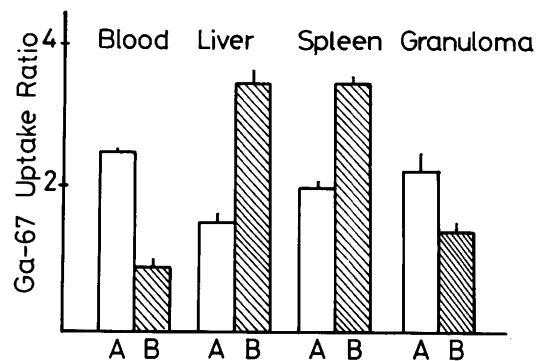


Fig. 3 Effect of the simultaneous administration of FeCl_3 on the uptake ratio of ^{67}Ga by several tissues. A: ^{67}Ga alone. B: ^{67}Ga was injected along with FeCl_3 . Each point represents the mean and SEM for five rats.

The effect of the simultaneous administration of FeCl₃ and ⁶⁷Ga on the uptake ratio of ⁶⁷Ga into inflammatory and normal tissues

The simultaneous administration of FeCl₃ and ⁶⁷Ga greatly increased the uptake ratios of ⁶⁷Ga into liver and spleen as compared with ⁶⁷Ga alone but decreased those into blood and granuloma (Fig. 3). The decrease in the uptake ratio of ⁶⁷Ga into blood by the simultaneous administration was, however, much smaller than that following the administration of FeCl₃ 5 min before and 30 min after the injection of ⁶⁷Ga.

DISCUSSION

We used ferric chloride as the iron reagent because ferric citrate poorly dissolved in saline, although Hayes et al.⁶ used ferric citrate to investigate the effect of iron on the distribution of ⁶⁷Ga. They have reported that ferric citrate slightly increased the blood distribution of ⁶⁷Ga. The results from Fig. 1 in the present study indicate, however, that FeCl₃ inhibits the binding of ⁶⁷Ga to transferrin, iron binding protein, in blood, so that the retention of ⁶⁷Ga in blood is greatly reduced by FeCl₃ and ⁶⁷Ga in an unbound form due to FeCl₃ is excreted in urine. Additionally FeCl₃ can decrease the uptake of ⁶⁷Ga by liver and spleen without disturbing the uptake of ⁶⁷Ga by the inflammatory lesions.

Hayes et al.⁶ reported that the uptake of ⁶⁷Ga by normal tissues such as liver or spleen occurred to a major extent by endocytosis in which transferrin is involved, and that the initial entry of ⁶⁷Ga into inflammatory lesions might occur in the same way as to that of normal soft tissues, while the uptake of ⁶⁷Ga by tumors occurs mainly by the diffusion of the unbound form. Vallabhajosula et al.⁷ reported that the uptake of ⁶⁷Ga into tumors was reduced by transferrin, as compared with ⁶⁷Ga-citrate alone, i.e., the uptake of ⁶⁷Ga by tumors occurred in an unbound form. In the present study, the results indicating that transferrin is involved in the uptake of ⁶⁷Ga by liver and spleen are consistent with those obtained by Hayes et al.,⁶ although the effect of the ferric citrate used by them on the liver and spleen was weaker than that of the ferric chloride used by us. We think, however, that the uptake of ⁶⁷Ga into inflammatory tissue occurs in an unbound form in the same way as into tumors. It should be noted, however, that the inflammatory lesions employed by Hayes et al. were abscesses but those employed by us were granuloma tissues, not containing an abscess. Oberhaensli et al.¹⁹ have also reported that iron decreased the distribution of ⁶⁷Ga into the liver and abscess. Both they and Hayes et al.⁶ injected iron simultaneously with ⁶⁷Ga. Their results are

quite the opposite of ours. In our results the simultaneous injection of iron increased the uptake of ⁶⁷Ga into liver and spleen. We cannot understand these differences. They must inject iron and ⁶⁷Ga by separate routes. Moreover, Hayes et al.⁶ have reported that iron increased the distribution of ⁶⁷Ga in blood, although that in liver was decreased with iron. These results are unreasonable. The results from the present study show that iron can decrease the amount of ⁶⁷Ga in blood but cannot remove the ⁶⁷Ga that has already entered the liver and spleen. Since almost all ⁶⁷Ga is, however, bound to transferrin in blood,^{8,9} the decrease in ⁶⁷Ga in blood due to FeCl₃ is the decrease in ⁶⁷Ga in a bound form. Therefore the decrease in the uptake of ⁶⁷Ga into liver or spleen when FeCl₃ was administered resulted from the decrease in ⁶⁷Ga binding to transferrin. On the other hand, FeCl₃ had little effect on the uptake of ⁶⁷Ga by granuloma, i.e., ⁶⁷Ga might accumulate in the inflammatory tissue either in a transferrin bound form or in an unbound form.

Many workers have expressed confidence that ⁶⁷Ga is exclusively bound to and transported to various tissues by transferrin in the blood. At the tumor or inflammatory site, ⁶⁷Ga is then dissociated from the transferrin complex. Vallabhajosula et al.^{16,18} reported that acidic pH at the tumor site might be one of the factors involved in ⁶⁷Ga localization in tumors. Moreover, reduction of the pH to 6.8–7.0, below that of normal tissue (7.4), might be due to the accumulation of lactic acid produced by anaerobic glycolysis. We think that the pH at the granuloma site must also be reduced by anaerobic glycolysis. We think that ⁶⁷Ga is transported to the inflammatory site either in a transferrin bound form or in an unbound form. Gallium-67 in an unbound form directly enters inflammatory tissue, whereas, ⁶⁷Ga in a bound form is dissociated under acidic conditions at the inflammatory site and then enters the inflammatory tissue. We conclude that transferrin is not involved in the uptake of ⁶⁷Ga into inflammatory tissue, such as granuloma, and that the uptake of ⁶⁷Ga into inflammatory tissue may be similar to that occurring in tumor tissues.

REFERENCES

1. Johnston GS: Clinical applications of gallium in oncology. *Int J Nucl Med Biol* 8: 249–255, 1981
2. Hoffer PB: Gallium and infection. *J Nucl Med* 21: 484–488, 1980
3. Hoffer PB: Use of gallium-67 for detection of inflammatory diseases: A brief review of mechanisms and clinical applications. *Int J Nucl Med Biol* 8: 243–247, 1981
4. Tzen KY, Oster ZH, Wabner HN, et al: Role of iron-binding proteins and enhanced capillary perme-

- ability on the accumulation of gallium-67. *J Nucl Med* 21: 31-35, 1980
5. Weiner R, Hoffer PB, Thakur ML: Lactoferrin: Its role as a Ga-67 binding protein in polymorphonuclear leukocytes. *J Nucl Med* 22: 32-37, 1981
 6. Hayes RL, Rafter JJ, Carlton JE, et al: Studies of the in vivo uptake of Ga-67 by an experimental abscess: Concise communication. *J Nucl Med* 23: 8-14, 1982
 7. Vallabhajosula SR, Goldsmith SJ, Lipszyc H, et al: ⁶⁷Ga-transferrin and ⁶⁷Ga-lactoferrin binding to tumor cells: Specific versus nonspecific glycoprotein-cell interaction. *Eur J Nucl Med* 8: 354-357, 1983
 8. Tsan MF, Scheffel U, Tzen KY, et al: Factors affecting the binding of gallium-67 in serum. *Int J Nucl Med Biol* 7: 270-273, 1980
 9. Vallabhajosula SR, Harwing JF, Siemsen JK, et al: Radiogallium localization in tumors: Blood binding and transport and the role of transferrin. *J Nucl Med* 21: 650-656, 1980
 10. Wong H, Turner UK, English D, et al: The role of transferrin in the vivo uptake of gallium-67 in a canine tumor. *Int J Nucl Med Biol* 7: 9-16, 1980
 11. Turner UK, Noujaim AA, Lentle BC, et al: The effects of differing gallium-transferrin-anion complexes on the tumor uptake of gallium-67. *Int J Nucl Med Biol* 8: 357-362, 1981
 12. Larson SM, Rasey JS, Allen DR, et al: Common pathway for tumor cell uptake of gallium-67 and iron-59 via a transferrin receptor. *J Nat Cancer Inst* 64: 41-53, 1980
 13. Larson SM, Grunbaum Z, Rasey JS: The role of transferrin in gallium uptake. *Int J Nucl Med Biol* 8: 257-266, 1981
 14. Hayes RL, Rafter JJ, Byrd BL, et al: Studies on the in vivo entry of Ga-67 into normal and malignant tissue. *J Nucl Med* 22: 325-332, 1981
 15. Hayes RL: The interaction of gallium with biological systems. *Int J Nucl Med Biol* 10: 257-261, 1983
 16. Vallabhajosula SR, Harwig JF, Wolf W: Effect of pH on tumor cell uptake of radiogallium *in vitro* and *in vivo*. *Eur J Nucl Med* 7: 462-468, 1982
 17. Ohkubo Y, Kohno H, Suzuki T, et al: Relation between ⁶⁷Ga uptake and the stage of inflammation induced by turpentine oil in rats. *Radioisotopes* 34: 7-10, 1985
 18. Vallabhajosula SR, Harwig JF, Wolf W: The mechanism of tumor localization of gallium-67 citrate: Role of transferrin binding and effect of tumor pH. *Int J Nucl Med Biol* 8: 363-370, 1981
 19. Oberhaensli RD, Mueller RM, Fridrich R: Different action of deferoxamine and iron on Ga-67 abscess detection in rats. *J Nucl Med* 25: 668-672, 1984