

patient (20 W.). The plasma renin substrate concentration ranged between 600 and 700 ng/ml in four patients with primary aldosteronism and in four with renal hypertension.

#### 2) Measurement of plasma renin concentration

Plasma renin substrate in the sample was destroyed and incubated with adequate amount of exogenous renin substrate (Skinner's method).

Three kinds of renin substrate were tested for this purpose. In human pool plasma and nephrectomised sheep plasma, there were substances which generated Angiotensin I after 4 to 6 hours incubation at 37°C, however after Haas' treatment, there was no Angiotensin I generation. In one mg of Cohn fraction IV-4 there were also Angiotensin I generating substances.

### Purification of $^{125}\text{I}$ -Glucagon by Microfine silica (Quso G-32)

Yoshinori ITO, Tetsuya OGASAWARA and Akira KIHARA

*First Department of Internal Medicine, Sapporo Medical College*

The  $^{125}\text{I}$ -glucagon labelling by chloramin T method showed a damage of the hormone which bound to plasma protein non-specifically and migrated with the protein on hydrodynamic flow electrophoresis. In an aim to purify the labelled glucagon, separation was carried out on sephadex G-10 column, and the purified fraction was mixed with 20 mg of Quso G-32 in 3 ml of 0.2 M glycine

buffer solution (pH 8.8).

The mixture was centrifuged and the precipitate was washed with 2 ml of the same buffer solution. The washed precipitate was then suspended in 2 ml of ethanol-0.5 N HCl (3:1, v/v), and was confirmed to be pure chromato-electrophoretically.

The purified  $^{125}\text{I}$ -glucagon reacted with a glucagon antibody (30K) satisfactorily.

### The Fundamental Study of Competitive Binding Radioassay for 25 OH D<sub>3</sub> in Human Plasma

S. DOKOH\*, M. FUKUNAGA\*, I. YAMAMOTO\*, K. TORIZUKA\* and R. MORITA\*\*

*\*Department of Radiology, Kyoto University Hospital, \*\*Central Clinical Radioisotope Division, Kyoto University Hospital*

The human plasma 25 Hydroxyvitamin D<sub>3</sub> (25 OH D<sub>3</sub>) levels were successfully measured by a competitive protein binding assay, using Vitamin D deficient rat serum as Vitamin D binding protein, for which a rat had been fed with Vitamin D, calcium, phosphorous deficient meals for 3 months. We have investigated some fundamental problems on performing this assay systems, 1: on the stability of 25 OH D<sub>3</sub> in ethanol and in serum under different conditions; (i.e. temperature, N<sub>2</sub>, O<sub>2</sub> and CO<sub>2</sub> gas, and ultraviolet irradiation) 2: on the effect of incubation time with binding protein, as well as incubation time with dextran coated charcoal (DCC), 3: variability of replacement curves obtained by changing the adding order of binding protein, radiolabeled and standard 25 OH D<sub>3</sub>, 4: recovery, and 5: cross reactivity with other Vitamin D analogues. 25 OH D<sub>3</sub> in ethanol was considerably destroyed by ultraviolet irradiation and by CO<sub>2</sub> gas atmosphere,

but was stable with temperature ranging from -20°C to 25°C under O<sub>2</sub> or N<sub>2</sub> gas as long as two days. Binding protein which was considered as Vitamin D transport protein had a high affinity to 25 OH D<sub>3</sub>, reaching maximum binding (Approximately 60%) within 5 minutes. The adsorptive reaction of DCC continued as long as 4 hours. Standard curves were variable by changing the mixing order of binding protein, radiolabeled and standard 25 OH D<sub>3</sub>, among those, the standard curve obtained by adding, : standard 25 OH D<sub>3</sub>, : radiolabeled 25 OH D<sub>3</sub>, and : binding protein; in this order, was the most practical and suitable for clinical use. Finally our assay system provided sensitivity of 0.1 ng/tube, excellent recovery (98.5±3.0%), poor cross-reactivity with other Vitamin D analogues such as D<sub>3</sub>, D<sub>2</sub>, 1-25 (OH)<sub>2</sub> D<sub>3</sub>, and 1α OH D<sub>3</sub>, and was proved to be useful for clinical application.